

## Epigenetics in allergy

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During the last two decades, great efforts have been focused on understanding genetic susceptibility to allergic diseases and have revealed that the diseases are polygenic and multifactorial. However, the genetic approaches could not fully explain heritability of allergic diseases (missing heritability), and the recent increase in incidence and prevalence of atopic diseases<sup>1,2)</sup>. The diseases are now thought to be greatly influenced by interactions between genetic predisposition and environmental factors, such as smoking, ozone, diesel exhaust particles (DEPs), SO<sub>2</sub> and NO<sub>2</sub><sup>3)</sup>. The environmental factors can cause epigenetic changes of individuals (or specific tissues and cell types)<sup>4)</sup>.

Epigenetics are defined as “stably heritable (and non-heritable) phenotype resulting from changes in a chromosome without alterations in the DNA sequence”<sup>5)</sup>, which include post-translational modification, DNA methylation, histone modifications and small non-coding RNAs. The epigenetic modifications, especially DNA methylation and histone modifications, plays key roles in T cell development<sup>6-8)</sup>. For example, Th1-specific genes such as TBX21 and IFNG are demethylated during the Th1 maturation, whereas those are methylated in Th2 or Th17 cell development. DNA methylation of IFNG is in turn preserved in Th2 and Th17 cells. In contrast, Th2-specific cytokines, IL4 and IL13, loci are hypomethylated in Th2 cells

As advances of microarray and sequencing technology, whole-genome epigenetic profiling (epigenome-wide association study; EWAS) has been widely used in epigenetic research. Microarrays have been the most frequently used methods in epigenomic study, with several platforms and protocols available for detecting DNA methylation<sup>9)</sup>. The microarrays have also been used to assess genome-wide histone modifications by using chromatin immunoprecipitation using specific antibodies for modified histone molecules, followed by hybridization on microarrays (ChIP-chip). Recently, next-generation sequencing has been applied to profiling DNA methylation (MBD-seq) and histone modifications (ChIP-seq)<sup>9)</sup>.

Many EWAS in allergic diseases have been performed, and some of them have reported differential methylation patterns between allergic patients and normal controls<sup>10)</sup>. Among allergic diseases, asthma has

been main target of EWAS for the last five years. Stefanowicz et al., tried to characterize differences in methylation within airway epithelial cells (AECs) and peripheral blood mononuclear cells (PBMCs) amongst healthy, atopic and asthmatic subjects using<sup>11</sup>. Although they identified 8 differentially methylated sites in AECs derived from asthmatics compared to atopics, there was no difference in the methylation status of PBMCs between disease phenotypes. Kim et al., in a study using bronchial mucosa tissues of atopic asthmatics, non-atopic asthmatics, and normal controls, reported 6 hypermethylated and 49 hypomethylated loci in the bronchial mucosa of atopic asthmatics compared to those of non-atopic asthmatics. However, the methylation levels in the mucosa of asthmatics and normal controls were similar<sup>12</sup>. Taken together, no DNA methylation changes were yet found to be directly associated with asthma in EWAS. In contrast, several EWAS demonstrated significantly different methylation patterns associated with asthma sub-phenotypes such as serum IgE levels<sup>13,14</sup>, prenatal tobacco smoke exposure<sup>15</sup>, pollutant exposure including diesel exhausted particles<sup>16-19</sup>, obesity<sup>20</sup>, and aspirin-hypersensitivity<sup>21</sup>. In fact, in our recent unpublished data, although peripheral CD4+ and CD8+ lymphocytes showed no differences between asthmatics and normal subjects, several loci showed different methylation levels in refractory asthmatics compared to those in controlled asthmatics. These results suggest that well-defined phenotypic characterization is important in epigenomic studies in asthma and in other allergic diseases, as well as improvement in sample size, replication efforts, and type of samples analyzed.

## References

1. Potaczek DP, Harb H, Michel S, Alhamwe BA, Renz H, Tost J. Epigenetics and allergy: from basic mechanisms to clinical applications. *Epigenomics*. 2017;9:539-71.
2. Renz H, Conrad M, Brand S, Teich R, Garn H, Pfefferle PI. Allergic diseases, gene-environment interactions. *Allergy*. 2011;66 Suppl 95:10-2.
3. von Mutius E. The environmental predictors of allergic disease. *J Allergy Clin Immunol*. 2000;105:9-19.
4. Harb H, Alashkar Alhamwe B, Garn H, Renz H, Potaczek DP. Recent developments in epigenetics of pediatric asthma. *Current opinion in pediatrics*. 2016;28:754-63.
5. Berger SL, Kouzarides T, Shiekhhattar R, Shilatifard A. An operational definition of epigenetics. *Genes & development*. 2009;23:781-3.
6. Aune TM, Collins PL, Chang S. Epigenetics and T helper 1 differentiation. *Immunology*. 2009;126:299-305.
7. Cohen CJ, Crome SQ, MacDonald KG, Dai EL, Mager DL, Levings MK. Human Th1 and Th17 cells exhibit epigenetic stability at signature cytokine and transcription factor loci. *Journal of immunology (Baltimore, Md : 1950)*. 2011;187:5615-26.
8. Tripathi SK, Lahesmaa R. Transcriptional and epigenetic regulation of T-helper lineage specification. *Immunol Rev*. 2014;261:62-83.
9. Yang IV, Schwartz DA. Epigenetic mechanisms and the development of asthma. *J Allergy Clin Immunol*. 2012;130:1243-55.
10. Nestor CE, Barrenas F, Wang H, Lentini A, Zhang H, Bruhn S, et al. DNA methylation changes separate allergic patients from healthy controls and may reflect altered CD4+ T-cell population structure. *PLoS Genet*. 2014;10:e1004059.

11. Stefanowicz D, Hackett TL, Garmaroudi FS, Gunther OP, Neumann S, Sutanto EN, et al. DNA methylation profiles of airway epithelial cells and PBMCs from healthy, atopic and asthmatic children. *PLoS One*. 2012;7:e44213.
12. Kim YJ, Park SW, Kim TH, Park JS, Cheong HS, Shin HD, et al. Genome-wide methylation profiling of the bronchial mucosa of asthmatics: relationship to atopy. *BMC Med Genet*. 2013;14:39.
13. Liang L, Willis-Owen SA, Laprise C, Wong KC, Davies GA, Hudson TJ, et al. An epigenome-wide association study of total serum immunoglobulin E concentration. *Nature*. 2015;520:670-4.
14. Yang IV, Pedersen BS, Liu A, O'Connor GT, Teach SJ, Kattan M, et al. DNA methylation and childhood asthma in the inner city. *J Allergy Clin Immunol*. 2015;136:69-80.
15. Breton CV, Siegmund KD, Joubert BR, Wang X, Qui W, Carey V, et al. Prenatal tobacco smoke exposure is associated with childhood DNA CpG methylation. *PLoS One*. 2014;9:e99716.
16. Clifford RL, Jones MJ, MacIsaac JL, McEwen LM, Goodman SJ, Mostafavi S, et al. Inhalation of diesel exhaust and allergen alters human bronchial epithelium DNA methylation. *J Allergy Clin Immunol*. 2017;139:112-21.
17. Jiang R, Jones MJ, Sava F, Kobor MS, Carlsten C. Short-term diesel exhaust inhalation in a controlled human crossover study is associated with changes in DNA methylation of circulating mononuclear cells in asthmatics. *Particle and fibre toxicology*. 2014;11:71.
18. Rossnerova A, Tulupova E, Tabashidze N, Schmuczerova J, Dostal M, Rossner P, Jr., et al. Factors affecting the 27K DNA methylation pattern in asthmatic and healthy children from locations with various environments. *Mutation research*. 2013;741-742:18-26.
19. Sominen HK, Zhang X, Biagini Myers JM, Kovacic MB, Ulm A, Jurcak N, et al. Ten-eleven translocation 1 (TET1) methylation is associated with childhood asthma and traffic-related air pollution. *J Allergy Clin Immunol*. 2016;137:797-805.e5.
20. Rastogi D, Suzuki M, Grealley JM. Differential epigenome-wide DNA methylation patterns in childhood obesity-associated asthma. *Scientific reports*. 2013;3:2164.
21. Cheong HS, Park SM, Kim MO, Park JS, Lee JY, Byun JY, et al. Genome-wide methylation profile of nasal polyps: relation to aspirin hypersensitivity in asthmatics. *Allergy*. 2011;66:637-44.